

Development of Enantioselective Synthetic Routes to (–)-Kinamycin F and (–)-Lomaiviticin Aglycon

Christina M. Woo, Shivajirao L. Gholap, Liang Lu, Miho Kaneko, Zhenwu Li, P. C. Ravikumar, and Seth B. Herzon*

Department of Chemistry, Yale University, New Haven, Connecticut 06520, United States

Supporting Information

ABSTRACT: The development of enantioselective synthetic routes to (-)-kinamycin F (9) and (-)-lomaiviticin aglycon (6) are described. The diazotetrahydrobenzo[*b*]fluorene (diazofluorene) functional group of the targets was prepared by fluoride-mediated coupling of a β -trimethylsi-lylmethyl- α , β -unsaturated ketone (38) with an oxidized naphthoquinone (19), palladium-catalyzed cyclization ($39 \rightarrow 37$), and diazo transfer ($37 \rightarrow 53$). The D-ring precursors **60** and **68** were prepared from *m*-cresol and 3-ethylphenol, respectively. Coupling of the β -trimethylsilylmethyl- α , β -unsaturated ketone **60** with the juglone derivative **61**, cyclization, and



diazo transfer provided the advanced diazofluorene 63, which was elaborated to (-)-kinamycin F (9) in three steps. The diazofluorene 87 was converted to the C_2 -symmetric lomaiviticin aglycon precursor 91 by enoxysilane formation and oxidative dimerization with manganese tris(hexafluoroacetylacetonate) (94, 26%). The stereochemical outcome in the coupling is attributed to the steric bias engendered by the mesityl acetal of 87 and contact ion pairing of the intermediates. The coupling product 91 was deprotected (*tert*-butylhydrogen peroxide, trifluoroacetic acid-dichloromethane) to form mixtures of the chain isomer of lomaiviticin aglycon 98 and the ring isomer 6. These mixtures converged on purification or standing to the ring isomer 6 (39–41% overall). The scope of the fluoride-mediated coupling process is delineated (nine products, average yield = 72%); a related enoxysilane quinonylation reaction is also described (10 products, average yield = 77%). We establish that dimeric diazofluorenes undergo hydrodediazotization 2-fold faster than related monomeric diazofluorenes. This enhanced reactivity may underlie the cytotoxic effects of (-)-lomaiviticin A (1). The simple diazofluorene 103 is a potent inhibitor of ovarian cancer stem cells (IC₅₀ = 500 nM).

INTRODUCTION

Lomaiviticins A–E (1–5, Figure 1) are the most complex members of a family of bacterial metabolites that contain a diazotetrahydrobenzo[b]fluorene (diazofluorene) functional group.¹ The lomaiviticins are structurally related to the monomeric diazofluorenes known as the kinamycins (7–10).² No less than 10 distinct kinamycins have been described; kinamycins A (7), C (8) and F (9), and the epoxykinamycin FL-120B' (10),³ are representative.

The kinamycins and lomaiviticins are potent antiproliferative and antimicrobial agents, and a large body of data now supports bioreductive activation⁴ of the diazofluorene as a first step in a cascade leading to cell death.⁵ It has been proposed that redox cycling of the quinone to produce reactive oxygen species (ROS),^{5a,g,j} formation of vinyl radical (11),^{5c-e,h,j} *o*-quinone methide (12),^{5c-e,h-j} and acylfulvene (13) intermediates by reduction,^{4a,51} or addition of biological nucleophiles to the diazo substituent (14),^{5b} may contribute to cytotoxicity (Figure 2). The targets of these metabolites are not known; various kinamycins and lomaiviticin A (1) are known to cleave or nick dsDNA in vitro^{1a,5h,j} and in tissue culture,^{5g} and a protein target has been implicated in the activity of kinamycin F (9).^{5k}

Synthetic planning toward these targets is significantly complicated by the sensitivity of the diazofluorene toward

nucleophiles and reducing agents. In addition, the β -alkoxyenone substructure of lomaiviticin A (1) is unstable toward an elimination–aromatization pathway (eq 1), which

limits the range of conditions that are compatible with latestage intermediates. Moreover, syntheses and installation of the carbohydrate residues of the lomaiviticins, in particular, the β linked 2,6-dideoxyaminoglycoside residues, are expected to be challenging. Finally, formation of the central and highly congested bridging carbon–carbon bond in the lomaiviticins constitutes perhaps the most significant obstacle toward their synthesis.

To date, several completed routes to the kinamycins⁶ and studies toward the lomaiviticins⁷ have been described. Our own efforts have resulted in an efficient synthesis of (-)-kinamycin F (9),^{6d} the first synthesis of (-)-lomaiviticin aglycon (6),⁷ⁱ and the development of a general and versatile strategy to prepare

Received: August 2, 2012 Published: October 3, 2012

Journal of the American Chemical Society

HO HO CH CH₃ CH ÓOH CHa R. R. kinamycin (-)-A (7) н Ar Ac Ac HO (-)-C (8) Ac Ac Ac (-)-F (9) CH CH₂-O HO N.N-dimethyl-L (-)-lomaiviticin aglycon (6) CH3 CH. (-)-lomaiviticin A (1) (-)-lomaiviticin B (2) (-)-FL-120B' (10) (-)-lomaiviticin C (3): R = R' = H (-)-lomaiviticin D (4): R = CH₃, R' = H and R = H, R' = CH₃ (-)-lomaiviticin E (5): R = R' = CH₃

Figure 1. Structures of (-)-lomaiviticins A-E (1-5, respectively), (-)-lomaiviticin aglycon (6), (-)-kinamycins A, C, and F (7-9, respectively), and the epoxykinamycin (-)-FL-120B' (10).



Figure 2. Reactive intermediates proposed to form from the kinamycins and lomaiviticins.

diazofluorenes of wide structural variability. We have discovered several new reactions during our investigations, have obtained insights into the factors influencing reductive activation of the diazofluorene, and have identified certain diazofluorenes as potent inhibitors of ovarian cancer stem cells. The full details of these investigations are described below.

RESULTS AND DISCUSSION

Development of a Convergent and Versatile Route to the Diazofluorene Function. Given the substantial body of evidence implicating the diazofluorene in the biological effects of the kinamycins and lomaiviticins,⁵ a versatile strategy to access this functional group was deemed valuable. We targeted diazofluorenes represented by the generalized structure **15** (Scheme 1). Deconstruction of **15** by excision of dinitrogen

Scheme 1. Strategy to Access the Diazofluorene Functional Group



and a one-carbon synthon, followed by separation of the naphthoquinone and enone residues, formed the hypothetical synthetic precursors 16 and 17. Initial efforts focusing on appending the one-carbon synthon and dinitrogen substituents to the naphthoquinone 16 were unsuccessful.⁸ Therefore, we pursued a strategy comprising addition of the one-carbon substituent to the enone, followed by bond formation to the quinone, as discussed below.

In preliminary studies, we employed the hydrazone 18 (Scheme 2), which was obtained in one step from cyclohex-2-

Article





ene-1-one.⁹ The hydrazone and enoxysilane functions of **18** were envisioned to serve as handles for bond formation to the naphthoquinone. We discovered that activation of **18** with tris(diethylamino)sulfonium difluorotrimethylsilicate [TASF-(Et)]¹⁰ in the presence of 5,8-dimethoxy-2,3-dibromonaphthoquinone (**19**)¹¹ cleanly formed the α -quinonylated product **20** in 73% yield as a single detectable *anti* diastereomer (¹H NMR analysis).

A broad range of enoxysilanes undergo α -quinonylation in high yield (Table 1). In cases where the enoxysilane possesses a β -stereocenter, the coupling products were formed with high anti selectivity (>5:1; entries 2, 3, 5, 7; 20, 64-86%). Unsymmetrical naphthoquinones couple with regiocontrol, provided that the reagent contains a substituent to differentiate the 2- and 3-positions (entries 4 and 5, 61 and 76%, respectively). Enoxysilanes derived from cyclopentanone (entry 6, 85%), acyclic ketones (entry 9, 82%), and heterocyclic ketones (entry 10, 85%) are also competent nucleophiles. α -Quaternary ketones are formed in good yield (entry 8, 65%). Common methods for the construction of carbon-carbon bonds to quinones include metal-catalyzed cross-coupling reactions,¹² lithiation of halogenated hydroquinone ethers,¹ and the oxidative addition of boronic acids.¹⁴ This fluoridemediated coupling reaction provides a useful complement to these protocols.

Synthesis of the quinonylated ketone 20 was encouraging since it contained all of the carbon atoms of our target. We envisioned forming the final ring by intramolecular 1,4-addition of the hydrazone to the naphthoquinone, followed by loss of hydrogen bromide $(20\rightarrow31$, Scheme 3). However, under a variety of conditions, the cyclization product 31 could not be formed (UPLC/MS analysis), potentially due to the *anti* arrangement of the quinone and hydrazone substituents of 20.⁸

Table 1. Scope of the α -Quinonylation Reaction^{*a*}



 $^a\mathrm{Conditions:}\ \mathrm{TASF(Et)}\ (1.10\ \mathrm{equiv}),\ \mathrm{CH}_2\mathrm{Cl}_2,\ -78\ ^\circ\mathrm{C}.\ ^b\mathrm{THF}\ \mathrm{used}\ \mathrm{as}$ solvent.

Scheme 3. Attempted Elaboration of the Hydrazone 20



We next targeted the β -methyl- α , β -unsaturated ketone 35 (Scheme 4). We envisioned forming an extended enolate of 35, followed by 5-*endo*-trig cyclization to construct the tetracycle 36. The precursor 34 was synthesized by copper-catalyzed 1,4-addition of methyl magnesium chloride to cyclohex-2-ene-1-one (32), trapping with chlorotrimethylsilane (see 33), and coupling with 5,8-dimethoxy-2,3-dibromonaphthoquinone (19) (80% overall). Although we were unable to oxidize 34 by several conventional methods, we discovered that thermoloysis of 34 under an atmosphere of dioxygen formed the oxidation product 35 (13%). Heating solutions of 35 in the presence of palladium(II) acetate, bis(diphenylphosphino)ferrocene, and

Scheme 4. Synthesis of the Tetracycle 37



potassium carbonate afforded the hydroxyfulvene 37 in 77% yield. Attempts to cyclize 35 using bases alone were unsuccessful.

The cyclization product 37, and structurally related intermediates (vide infra), exist exclusively in the hydroxyfulvene (e.g. 37) rather than the keto tautomer (36, Scheme 4). Substituent-dependent ketone-hydroxyfulvene equilibria have been reported for related structures.^{Si} In the present system, the hydroxyfulvene tautomer may be stabilized by intramolecular hydrogen-bonding and π -conjugation of the hydroxy group with the carbonyl substituents.

Although we had made substantial progress toward our target, the low efficiency of the oxidation step $(34\rightarrow35)$ led us to reorganize the sequence of bond-forming events. Three considerations were critical in the development of our successful approach. First, we recognized that reoxidation (e.g., palladium acetate¹⁵ or benzeneselenyl chloride–hydrogen peroxide¹⁶) of conjugate addition products such as 33 provides a route to β -substituted- α,β -unsaturated ketones that circumvents the difficult oxidation observed above $(34\rightarrow35)$. Second, the fluoride-mediated quinonylation reaction appeared to be unique in its ability to unite the enone and quinone fragments, and we sought to retain this chemistry. Finally, although not mechanistically well-understood, the palladium-catalyzed cyclization $(35\rightarrow37)$ provided an efficient method to forge the cyclopentadiene ring of the target.

With these considerations in mind, the β -trimethylsilylmethyl- α , β -unsaturated ketone **38** was synthesized by coppercatalyzed 1,4-addition of trimethylsilylmethylmagnesium chloride to **32**, trapping with chlorotrimethylsilane, and reoxidation (76% overall, Scheme 5). TASF(Et)-mediated coupling of **38** with 2,3-dibromo-5,8-dimethoxynaphthoquinone (**19**) proceeded smoothly, to afford the ketoquinone product **39** (85%). The observed preference for γ -quinonylation may be steric in origin.

Scheme 5. Synthesis of the Ketoquinone 39



dx.doi.org/10.1021/ja307497h | J. Am. Chem. Soc. 2012, 134, 17262-17273

This γ -quinonylation reaction is also relatively general (Table 2). A variety of β -trimethylsilylmethyl- α , β -unsaturated ketones,



^aConditions: TASF(Et) (1.1 equiv), CH₂Cl₂, -78 °C. ^bTBAT (1.1-1.5 equiv), CH₂Cl₂, 0 °C. ^cNone detected by ¹H NMR analysis of the unpurified reaction mixture.

including those with quaternary carbon centers (entries 1, 2, 7, and 8, 57–96%), acetal (entries 3 and 4, 83 and 85%), *p*methoxybenzyl ether (entries 6 and 7, 99 and 65%), and ester substituents (entry 8, 96%), couple in high yields. As in our α quinonylation reaction, unsymmetrical, electronically biased naphthoquinones react with regiocontrol (entry 4, 85%). 3-(Trimethylsilylmethyl)cyclopent-2-ene-1-one is also a competent nucleophile for this reaction (entry 5, 67%). Nucleophiles bearing α -substitution, however, couple in diminished yields (entry 9, 22%), and the presence of a ketone substituent appears to be essential to achieve activation of the allylsilane function (entry 10). We have recently found that the benchstable, commercially available salt tetrabutylammonium difluorotriphenylsilicate (TBAT)¹⁷ may be employed in place of TASF(Et) (entries 1 and 3).

With the ketoquinone **39** in hand, we investigated methods to forge the final carbon-carbon bond of the diazofluorene $(39\rightarrow 37)$. Surprisingly, the conditions employed for the

cyclization of 35 were ineffective (entry 1, Table 3). Further optimization was conducted with stoichiometric palladium to facilitate accurate measurement. Among a variety of ligands and palladium precursors examined, the combination of palladium-(II) acetate and triphenylphosphine emerged as the most promising. Using these reagents and cesium carbonate as base, the hydroxyfulvene 37 was isolated in 31% yield (entry 2). Triethylamine (TEA, entry 3) and silver phosphate (entry 4) were less effective. By employing silver carbonate as base, raising the temperature to 80 °C, and using polymer-supported triphenylphosphine (PS-PPh₃, to simplify purification), the isolated yield of 37 was increased to 40% (entry 6). The low isolated yield of 37 was attributed to formation of the palladium complex 52, identified by UPLC/MS analysis and accounting for a significant mole fraction of 37 at these high loadings of palladium. Complexes between the structurally related anthracyclines and palladium are known.¹⁸ Although we were unable to ascertain the nature of the bonding interactions in 52 (e.g., η^5 , κ^2), attempts to displace the product 37 by addition of excess acetylacetone or bis(diphenylphosphino)ferrocene were unsuccessful. The substrate 50, which bears a quaternary center on the D-ring, formed the cyclization product 51 in 79% yield (entry 7), and a complex with palladium could not be detected during the reaction (UPLC/MS analysis). Using this substrate, we were also able to decrease the loading of palladium to 25 mol % (86% yield of 51, entry 8). In general, cyclizations of substrates bearing a quaternary center on the D-ring, including those employed en route to the targets (vide infra), are much more efficient than simple model systems such as 39.

Diazo transfer to the hydroxyfulvene 37 was all that was required to complete the synthesis of the diazofluorene. Historically, an early application of p-toluenesulfonyl azide $(p-TsN_3)$, by Doering,¹⁹ was in the synthesis of diazocyclopentadiene itself from lithium cyclopentadienide. The conjugate base of 37 possesses five inequivalent cyclopentadienide carbon atoms, but only addition to the desired position generates a stable diazo adduct. Exposure of the hydroxyfulvene 37 to triethylamine and common diazo transfer agents, such as methanesulfonyl azide (MSA),²⁰ p-acetamidobenzenesulfonyl azide (p-ABSA),²¹ or p-TsN₃,¹⁹ afforded low yields of the diazofluorene product 53 (Table 4, entries 1-3, 4-20%). Use of the more electrophilic imidazole-1-sulfonyl azide hydrochloride (ISA)²² formed the diazofluorene 53 in 74% yield in less than 1 h at 0 °C (entry 4). Trifluoromethanesulfonyl azide $(TfN_3)^{23}$ and nonaflyl azide²⁴ were slightly more effective (81 and 85%, respectively, entries 5 and 6). The latter is preferred due to its increased thermal stability and lower volatility.

The chemistry outlined above has allowed us to prepare a large variety of monomeric diazofluorenes. The antiproliferative activity of many of these compounds has been evaluated, and certain synthetic diazofluorenes have emerged as potent inhibitors of cancer stem cells (vide infra). Of greatest significance, these studies laid the foundation for syntheses of (-)-kinamycin F (9) and (-)-lomaiviticin aglycon (6), as described below.

Syntheses of the Enone Fragments. Our route to the diazofluorene defined the enone fragments required for syntheses of kinamycins and lomaiviticins, and these were prepared by the routes outlined in Scheme 6. Beginning with *m*-cresol (kinamycin series), silyl ether formation (triisopropylsilyl chloride, imidazole) and Birch reduction (lithium, ammonia) formed the cyclohexadiene **54** (99%, two steps). Regio- and stereoselective asymmetric dihydroxylation²⁵ then

Table 3. Optimization of the Palladium-Mediated Cyclization Reaction^a

		CH ₃ O O CH ₃ O O CH ₃ O O	Pd(OAc) ₂	CH ₃ O O CH ₃ O O CH ₃ O O H O			
		39 R = H 50 R = CH ₃		37 R = H 51 R = CH ₃	•PdPPh ₃ 52		
entry	substrate	solvent	mol % Pd	ligand	base	<i>T</i> (°C)	yield $(\%)^b$
1	39	DMF	40	dppf	K ₂ CO ₃	50	<5
2	39	THF	80	PPh_3	Cs_2CO_3	50	31
3	39	THF	100	PPh ₃	TEA	50	<5
4	39	THF	100	PPh ₃	Ag ₃ PO ₄	50	dec. ^c
5	39	PhCH ₃	75	PS-PPh ₃	Ag ₂ CO ₃	80	22
6	39	PhCH ₃	100	PS-PPh ₃	Ag ₂ CO ₃	80	40
7	50	PhCH ₃	100	PS-PPh ₃	Ag ₂ CO ₃	80	79
8	50	PhCH ₃	25	PS-PPh ₃	Ag_2CO_3	80	86

^{*a*}Conditions: 1–2 equiv of ligand (with respect to palladium) and 2–3 equiv of base (with respect to substrate) were employed. ^{*b*}Isolated yield after purification by flash-column chromatography. ^{*c*}Unidentified decomposition products were formed.

Table 4. Diazo Transfer to the Hydroxyfulvene 37^{a}

CH ₃ O C CH ₃ O C	diazo transfer Et ₃ N OH OH 37	r agent	CH30 CH30	$\overbrace{53}^{O} \xrightarrow{N_2}$
entry	diazo transfer agent (equiv)	<i>t</i> (h)	T (°C)	yield $(\%)^b$
1	MSA (7.5)	4	24	4
2	<i>p</i> -ABSA (5.0)	1.5	24	20
3	p-TsN ₃ (7.5)	4	24	18
4	ISA (2.5)	0.3	0	74
5	TfN_{3} (2.5)	0.3	24	81
6	$CF_3(CF_2)_3SO_2N_3$ (2.5)	0.3	0	85

^{*a*}Conditions: diazo transfer reagent (2.5–7.5 equiv), TEA (5.0–15 equiv), CH₃CN. ^{*b*}Isolated yield after purification by flash-column chromatography.



Scheme 6. Syntheses of the Enone 60

generated the vicinal diol **56** in 66% ee (55% yield). The use of the triisopropylsilyl protecting group was critical to the success of this step; presumably, its steric bulk prevents entry of the enoxysilane into the binding pocket of cinchona alkaloid– osmium complex, allowing selective oxidation of the less reactive alkene. The modest enantiomeric excess observed in this transformation was attributed to the negligible steric requirements of the methyl substituent, which renders discrimination of the faces of the alkene difficult. No attempt to optimize the enantiomeric excess was made, as we found that subsequent intermediates were highly crystalline and could be resolved to 97% ee.

The diol **56** was then transformed to the acetonide **58** by stirring with 2,2-dimethoxypropane (2,2-DMP) and pyridinium *p*-toluenesulfonate (PPTS, 88%). Treatment of **58** with phenylselenium chloride formed the α -selenyl ketone **59** (69%, >20:1 mixture of diastereomers). Finally, oxidation of **59** (hydrogen peroxide, pyridine) and thermal elimination of the resulting selenyl oxide formed an enone (not shown) that was homologated by copper-catalyzed 1,4-addition of trime-thylsilyl methylmagnesium chloride, trapping with chloro trimethylsilane, and reoxidation (**60**, 73% over two steps).

In the lomaiviticin series, silvlation and Birch reduction of 3ethylphenol provided the diene 55 (>99%). Asymmetric dihydroxylation of 55 formed the diol 57 (62%, 91% ee). Derivatives of 57 were used in our lomaiviticin studies (vide infra).

Completion of (-)-Kinamycin F (9). To complete the synthesis of (-)-kinamycin F (9), a suitable naphthoquinone coupling partner, biased toward addition at the C-3 position, was required.⁸ Toward this end, O-(methoxymethyl)-2-bromo-3-methoxyjuglone (61) was prepared by heating a methanolic solution of O-(methoxymethyl)-2,3-dibromojuglone and sodium carbonate (96%).8 Fluoride-mediated coupling with the β -trimethylsilylmethyl- α , β -unsaturated ketone **60** proceeded with complete regioselectivity, forming the C-3 substituted product 62 in 79% yield (Scheme 7). The cyclization and diazotransfer steps proceeded smoothly, as in our model system, to form the diazofluorene 63 (65%, two steps). To complete the synthesis we needed to develop a method for the stereocontrolled conversion of the ketone function of 63 to a trans-1,2-diol. Site-selectivity in the reduction step was a serious concern, as an earlier study by Feldman and Eastman^{5c,d} had established the diazofluorene as highly reactive toward reducing agents. To control selectivity, we pursued an α -oxidation, directed reduction sequence. Treatment of the diazofluorene 63 with triisopropylsilyl trifluoromethanesulfonate (TIPSOTf) and diisopropylethylamine (DIPEA) formed an enoxysilane (not shown) that was exposed to an excess of dimethyldioxirane in a mixture of methylene chloride and methanol at -40 °C. Under these conditions, the free α -hydroxy ketone 65 was obtained, presumably formed by in situ methanolysis of the silyloxy epoxide 64. The stereoselectivity of this oxidation [verified by

Scheme 7. Completion of the Synthesis of (-)-Kinamycin F (9)



successful conversion of **65** to (-)-kinamycin F (9)] is consistent with faster addition of the dioxirane to the convex face of the *cis*-fused 6-5 system. The α -hydroxyl group of **65** was then used to control selectivity in the reduction step.

We considered several reagents known to effect the directed reduction of α - and β -hydroxy ketones. This reaction is most commonly accomplished with tetramethylammonium triacetoxyborohydride,²⁶ but the use of acetic acid as cosolvent was undesirable. We employed borane-tetrahydrofuran complex, as this reagent had been shown to effect the directed reduction of α -hydroxy ketones, though the stereoselectivity for simple cycloalkanones was low.²⁷ To implement the reduction, the α hydroxyketone 65 was dissolved in tetrahydrofuran and cooled to -78 °C. Borane-tetrahydrofuran complex (1 equiv) was added, and the resulting mixture was warmed to -20 °C. Under these conditions the trans-1,2-diol 67 was formed in 58% yield. Minor amounts (<5%) of quinone reduction products were detected by UPLC/MS analysis, suggesting that the reduction proceeds via the in situ generated borinite ester 66. To complete the synthesis, the diol 67 was deprotected by treatment with hydrochloric acid in methanol at low temperature, to provide (-)-kinamycin F (9) in 65% yield.

Synthesis of (-)-Lomaiviticin Aglycon (6). With a route to the kinamycins established, we focused on the preparation of (-)-lomaiviticin aglycon (6). When we began our studies we hypothesized that, given the existence of the kinamycins, nature may synthesize lomaiviticins A (1) and B (2) by oxidative coupling of monomeric diazofluorenes (see Supporting Information for a working mechanism of the putative biosynthetic dimerization). Accordingly, we considered several synthetic methods to effect this transformation. Direct oxidative coupling of cycloalkanones,²⁸ oxidative coupling of ketone, ester, and amide enolates,²⁹ and single electron oxidation of enoxysilane³⁰ or enamine³¹ derivatives all had varying degrees of precedent. Two considerations led us to focus on enoxysilane coupling pathways. First, in our kinamycin work, we discovered that β -oxygenated enoxysilanes (Scheme 7) derived from protected diazofluorenes could be generated in near quantitative yield. Second, we expected that β -elimination would be problematic using enolate-based methods.

In a model study, we found that enoxysilanes derived from simple diazofluorenes could be oxidatively coupled in good yield.⁸ This motivated us to study this transformation in more detail using a fully functionalized substrate, and the diazofluorene 71 was prepared by the sequence outlined in Scheme 8. Oxidation¹⁵ of the enoxysilane 57 formed the enone 68 (92%). Treatment of the enone 68 with mesitylaldehyde

Scheme 8. Synthesis and Dimerization of the Bis(methoxymethyl)-endo-mesityldiazofluorene 71



dimethyl acetal³² in the presence of PPTS as catalyst formed the acetal 69 (12:1 mixture of endo and exo isomers, 80%). Homologation, fluoride-mediated coupling with 5,8-bis-(methoxymethyl)-2,3-dibromonaphthoquinone (70), cyclization, and diazo transfer provided the endo-mesityl diazofluorene 71 in 37% overall yield. Treatment of the endo-mesityl diazofluorene 71 with p-toluenesulfonic acid in methanol formed the (-)-monomeric lomaiviticin aglycon (72, 55%).³³ Alternatively, treatment of 71 with tert-butyldimethylsilyl trifluoromethanesulfonate (TBSOTf) and DIPEA formed an enoxysilane (not shown) that was oxidatively coupled (ceric ammonium nitrate, CAN) to provide the C_2 -symmetric dimeric product 73 (42%), as well as the monomeric quinone 74 (8%). In these experiments, only a single dimeric product was formed, as determined by ¹H NMR and UPLC/MS analysis of the unpurified reaction mixture. Although the C_2 -symmetry of the product was evident by NMR analysis, this element of symmetry also prevented rigorous assignment of relative stereochemistry. To address this, we deprotected the dimer 73 by heating with 20% trifluoroacetic acid in methanol at 50 °C. Under these strongly acidic conditions, the starting material was converted to the aglycon 75 (41%). This compound was completely characterized by multidimensional NMR analysis, and these data revealed the presence of the C-1/1' carbonyl substituents, providing evidence that ring closure to form the hemiketal structure of lomaiviticin B (2) had not occurred. Based on this observation, and our kinamycin studies (vide supra), we assigned the stereochemistry of 75 as the unnatural anti,anti-configuration. In retrospect, the formation of the anti, anti-diastereomer 73 is not surprising; the concave topology of the molecule and the steric congestion introduced by the mesityl protecting groups both favor bond formation to the α -face of the monomer 71.

Aromatization issues notwithstanding, among several strategies considered to arrive at the natural *syn,syn*-configuration, double-epimerization of the *anti,anti*-aglycon 75 seemed the most straightforward. An extensive battery of conditions were evaluated with the goal of enolizing the ketone functional groups within 75. We hypothesized that even if the relative energies of the diastereomers were similar, cyclization to form lomaiviticin aglycon (6) should drive the equilibrium forward. Unfortunately, however, the *anti,anti*-aglycon 75 exhibited limited stability toward nucleophiles and bases. Under forcing acidic conditions, 75 was remarkably inert. For example, no deuterium was detected in the α -positions of 75 after heating with trifluoroacetic acid (20% v/v) in methanol- d_4 for 7 min at 120 °C under microwave irradiation (¹H NMR analysis).

We then turned our attention toward epimerization of the protected *anti,anti*-dimer 73. In this case, hemiketal formation is not possible, but we expected that protonation of an enol(ate) from the convex face to afford the desired *syn,syn*-stereochemistry would be kinetically favored. Although this protected intermediate was significantly more robust, activation of the α -protons was not possible. This may be due to the increased steric bulk about these protons, originating from the mesityl acetal protecting groups. After several months of focused experimentation, our epimerization studies were abandoned.

We next explored the installation of a removable functional group at the α -position of the diazofluorene (Scheme 9). In this approach, the stereochemistry of the resulting dimer would be

Scheme 9. Synthesis and Reactions of α -Functionalized Diazofluorenes 77



ultimately controlled by removal of this functional group, rather than the dimerization step. Several α -functionalized diazofluorenes were prepared from the enoxysilane 76, including the α bromo, α -fluoro, α -phenylselenyl, and α -phenylsulfenyl ketones 77a-d, respectively. However, with the exception of the α fluoroketone derivative 77b, all attempts to generate α substituted enoxysilanes from these functionalized diazofluorenes were unsuccessful. Although conditions for the reductive cleavage of α -fluoroketones are not widespread,³⁴ we attempted to advance the fluorinated product 77b. Treatment of 77b with TBSOTf and TEA formed the enoxysilane 78. Exposure to CAN and sodium bicarbonate formed a single C_1 -symmetric dimeric product (79, 39%). The presence of extensive oxidation and fluorine atoms about the newly-formed bond made complete characterization of 79 difficult, but all NMR data are consistent with carbon-oxygen bond formation (as shown), rather than the expected carbon-carbon coupling product.

Although we were cognizant of the reactivity of the diazofluorene toward reducing agents, ^{5c,d,h,i} we prepared⁸ the α -phenylselenyl and α,β -epoxy ketones **80** and **81** and examined the possibility of conducting a reductive coupling reaction by formation of an α -keto radical (Figure 3).



Figure 3. Structures of the α -phenylselenyldiazofluorene **80**, the α , β -epoxydiazofluorene **81**, and the monomeric lomaiviticin aglycon (72).

Unfortunately, under a variety of reducing conditions⁸ extensive decomposition of the substrates was observed. We also attempted the direct oxidative coupling of the monomeric lomaiviticin aglycon (72). The flexibility introduced by removal of the cyclic protecting group was anticipated to allow access to the natural *syn,syn*-stereochemistry. However, efforts to dimerize 72, directly or in the presence of air or added oxidants [CAN, bis(trifluoroacetoxy)iodobenzene, potassium ferricyanide], were unsuccessful. In some instances oxidation to a putative diquinone (UPLC/MS and NMR analysis, not shown) was observed, but this could not be manipulated to a dimeric product.

The extensive studies described above served to define the stability profile of synthetic diazofluorenes: while they were reactive toward reducing agents and nucleophiles, they were robust toward strong oxidants. With this in mind, we prepared the C-3-deoxydiazofluorene **83** (Scheme 10) because we believed that removal of the C-3 oxygen substituent would bias the system for oxidative coupling *anti* to the ethyl substituent, to provide the natural relative configuration. Stereoretentive oxidation of the C-3/3' C–H bonds of the product would then provide lomaiviticin aglycon (**6**). This strategy found encouraging precedent in the work of Wender and Baran, who had demonstrated that highly selective C–H hydroxylations can be effected in complex settings using dioxirane-based reagents.³⁵

The synthesis of the protected C-3/3'-dideoxydimer **84** began with the ketone **82**.³⁶ Protection of the secondary alcohol (chloromethyl methyl ether, DIPEA) and elaboration as before provided the C-3-deoxydiazofluorene **83** (24% over five



steps). Enoxysilane formation (TBSOTf, DIPEA), followed by addition of CAN, formed the oxidative coupling product **84** in 60% yield as a single detectable diastereomer. The relative stereochemistry of **84** was confirmed by observation of a W-plane coupling between the C-2 and C-4 hydrogen atoms. Deprotection of **84** (boron tribromide, dichloromethane) formed C-3/3'-dideoxylomaiviticin aglycon (**85**, 46%). Unfortunately, neither **84** nor **85** were reactive toward dimethyldioxirane or trifluorodimethyldioxirane, perhaps due to the steric congestion about the targeted C–H bonds.

At this juncture two invariable trends dominated our synthetic planning. First, single electron oxidation of monomeric enoxysilanes was the only method that provided dimeric products. Second, modification of the stereochemistry following the dimerization appeared unachievable. These considerations led us to focus on the synthesis of substrates that might be compatible with the enoxysilane coupling protocol and also provide the natural stereochemistry on dimerization. We considered the use of acyclic protecting groups on the vicinal diol function; however, intermediates bearing acyclic alkyl or silyl ether protecting groups were less stable toward elimination of the β -oxygen substituent. The limited stability of the diol **68** prevented installation of smaller (unactivated) cyclic protecting groups, such as formaldehyde acetal.

Inspection of molecular models, and later, computational studies⁸ clearly revealed the origins of the steric bias in the dimerization of the endo-mesityl diazofluorene 71. In the energy-minimized structure,⁸ the mesityl substituent protrudes into the cavity of the 6-5 system. Approach from the α -face, to afford the undesired anti, anti-stereochemistry, is kinetically favored. In considering ways to invert this bias, we recognized that epimerization of the mesityl substituent, to the exo orientation, might promote bond formation to the desired (concave) face of the enoxysilane. The exo-mesityl diazofluorene 87 was prepared by a modification of our route to the endoisomer 71 (Scheme 11). Under conditions developed for protection of the diol 68, the endo-acetal is the kinetically favored product. Warming of the reaction mixture to 50 °C promotes epimerization of the acetal, to form a 1:1 mixture of endo and exo diastereomers (86). Resubjection of the diastereomerically pure exo isomer (obtained by careful purification on ca. 25-mg scales) formed a 1:0.8 mixture of exo:endo isomers, suggesting they are nearly equal in energies. In practice, the mixture of endo and exo acetals 86 (and





subsequent intermediates) was advanced through the diazo transfer step, at which point the *endo-* and *exo-*diazofluorenes 71 and 87 were isolated separately.

Ultimately, the diazofluorene 87 was successfully advanced to lomaiviticin aglycon (6) by a sequence comprising enoxysilane formation, oxidative dimerization, and deprotection (Scheme 12). Among a large number of oxidants evaluated for their

Scheme 12. Products Identified in the Oxidative Coupling of the *exo*-Mesityl Diazofluorene 87



ability to dimerize enoxysilanes derived from 87, only two, CAN and manganese tris(hexafluoroacetylacetonate) (94),^{37,38} were effective [oxidation potentials (vs Fc) = 0.88 and 0.9 for CAN³⁹ and 94,³⁸ respectively]. The diazofluorene 87 and up to five products are obtained in these transformations: the undesired *anti,anti* and *syn,anti* dimers 90 and 92, respectively, the desired *syn,syn*-dimer 91, the aromatized–oxidized monomer 74, and the α -nitrodiazofluorene 93 (when CAN was employed).

A detailed evaluation of the oxidative dimerization has revealed that the product distribution, efficiency, and stereoselectivity of the transformation are dependent upon enoxysilane structure (silicon substituents), choice of oxidant, and solvent. Selected experiments that provide an overall representation of the factors influencing this reaction are shown in Table 5. Treatment of the *tert*-butyldimethylsilyl derivative

Table 5. Dimerization of the exo-Mesityl Diazofluorene 87^{a}

					yield (%)			
entry	R_3^{b}	oxidant	solvent	90	91	92	74	87
$1^{c,d}$	88	CAN	CH ₃ CN	24	nd ^e	nd	nd	3
2^{f}	88	94	PhH	<5	nd	nd	28	5
3^g	89	94	CH_2Cl_2	26	nd	<5	15	<5
4 ^{<i>c</i>}	89	CAN	CH ₃ CN	17	6	10	8	8
5f	89	94	PhH	12	26	<5	15	36

^{*a*}Isolated yields after purification by preparatory thin-layer chromatography. ^{*b*}**88**, R₃ = *t*-Bu(CH₃)₂; **89**, R₃ = (CH₃)₃. ^{*c*}CAN (2.0 equiv), sodium bicarbonate (20 equiv), CH₃CN, -35 °C. ^{*d*}The α -nitrodiazofluorene **93** was obtained in 14% yield. ^{*e*}None detected. ^{*f*}Mn(hfacac)₃ (**94**, 1.20 equiv), PhH, 24 °C. ^{*g*}Mn(hfacac)₃ (**94**, 1.20 equiv), CH₂Cl₂, 24 °C.

88 with ceric ammonium nitrate afforded a 24% yield of the undesired anti, anti-diastereomer 90, 14% of the α -nitrodiazofluorene 93, and extensive amounts of unidentified decomposition products (entry 1). Within the limits of UPLC/MS analysis, no additional dimeric products were formed. Oxidation of 88 with manganese tris(hexafluoroacetylacetonate) (94) in benzene resulted in aromatization (74, 28%) and production of a large amount of unidentified decomposition products; less than 5% of the anti, anti diastereomer 90 was formed (entry 2). Oxidation of the trimethylsilyl derivative 89 was much more efficient using 94, and the undesired anti,anti diastereomer 90 was obtained in 26% yield when dichloromethane was employed as solvent (entry 3). Encouragingly, oxidation of 89 with CAN formed the three dimeric products 90-92 in 17, 6, and 10% yield, respectively, as well as the aromatized monomer 74 (8%) and the diazofluorene 87 (8%, entry 4). By employing the trimethylenoxysilane 89, manganese tris(hexafluoroacetylacetonate) (94) as oxidant, and benzene as solvent, the desired syn,syn coupling product 91 was obtained as the major product (26%, entry 5).

Smaller amounts of the undesired *anti,anti-*coupling product **90** (12%) were also isolated, and only trace quantities of the *syn,anti-*dimer **92** could be detected under these conditions (UPLC/MS analysis). These latter conditions were reproducibly executed on 100-mg scales.

The complexities of radical cation reactions⁴⁰ and low mass balances in Table 5 caution against overinterpretation of these results. However, the enhanced anti, anti stereoselectivity in the dimerization of 89 using manganese tris-(hexafluoroacetvlacetonate) (94) in benzene (entry 5) compared to dichloromethane (entry 3) may be due to formation of a tight ion pair in which the manganese anion is associated with the convex face of the radical cation.⁸ Kochi has shown that ion pairs formed from enoxysilanes are closely associated in nonpolar solvents.⁴¹ This association may direct bond formation to the more hindered (concave) face of the substrate. Under identical conditions, oxidation of the trimethylenoxysilane derived from the endo-isomer 71 provides the anti,anti-dimer 73 exclusively (38%),8 suggesting that shielding by the endo-mesityl substituent overrides the steric encumbrance of the manganese anion.

The final obstacle involved deprotection of the syn,syn-dimer 91. While the anti,anti-dimeric products 73 and 90 were stable toward strongly acidic conditions (vide supra; see Supporting Information for deprotection of 90), the syn,syn-dimer 91 decomposed under mildly acidic conditions. We attribute the heightened reactivity of 91 to the anti arrangement of the α proton and β -oxygen substituents, which facilitates β elimination. After many experiments that resulted in unidentifiable decomposition products, a useful lead was obtained. Treatment of 91 with excess trifluoroacetic acid in wet dichloromethane at -35 °C resulted in rapid formation of the mono mesityl-protected aglycon 95 (UPLC/MS analysis), which slowly converted to the aromatized aglycon 97 (Scheme 13A). We reasoned that the aromatized product 97 was forming by elimination of the β -oxocarbenium ion 96. To prevent this, we conducted the deprotection in the presence of a large excess of *tert*-butyl hydroperoxide (Scheme 13B).⁴² LC/ MS analysis revealed that 95 accumulated under these conditions as well, but now neutralization of the reaction mixture at low temperature (-78 °C) followed by extractive workup provided mixtures of the chain isomer of lomaiviticin aglycon 98 as well as the expected ring isomer 6 (98:6 = 8-2:1), rather than the dehydrated product 97. Presumably, under





these conditions trapping of the oxocarbenium ion **96** by the peroxide nucleophile is faster than the rate of β -elimination.

The chain isomer **98** was characterized by multidimensional NMR analysis, which established the presence of the C-1 carbonyl substituent. Attempted separation of this mixture by preparative thin-layer chromatography resulted in cyclization to form the ring isomer **6** exclusively (39–41% overall).

The aglycon 6 was fully characterized by NMR, IR, and HRMS analysis and all data are consistent with the structure shown. However, we observed some differences in NMR chemical shifts between the bis(hemiketal) core of 6 and those of lomaiviticin B (2). In particular, the H-2 and C-2 resonances of 6 are observed at 3.41 and 62.1 ppm, respectively (30% DMF- d_7 -CD₃OD), while those of lomaiviticin B (2) are reported to resonate at 2.64 and 44.0 ppm, respectively (CD₃OD; the aglycon 6 is sparingly soluble in CD₃OD, which prohibited acquisition of spectroscopic data in this solvent).^{1a} The C-2 and H-2 resonances of lomaiviticin aglycon 6 are in good agreement with a model system prepared by Nicolaou and co-workers (δ H-2 = 2.87, δ C-2 = 60.4, C₆D₆),^{7a} and NMR spectroscopic data for the chain isomer 98 matched closely those of lomaiviticin A(1). To provide further evidence for connectivity and stereochemistry in 4, we isolated the monomesityl aglycon 95 (Scheme 13A) by arresting the deprotection of 91 prematurely. The ¹H and ¹³C resonances within the bis(cyclohexenone) substructure of 95 were resolved in CDCl₃ and were assigned by multidimensional NMR analysis, providing confirmation of connectivity. An NOE between H-4' and H-2' was observed, providing further confirmation of the syn,syn-stereochemistry of the dimerization product 91 (Scheme 13C).

The data above left little doubt as to the structure of synthetic lomaiviticin aglycon (6). We recognized, however, that natural lomaiviticin B (2) was obtained as its bis-(trifluoroacetate) salt, following a final HPLC purification step.^{1a} To probe for potential effects of charge on chemical shifts, lomaiviticin B (2) was obtained by partial hydrolysis of lomaiviticin A^{1b} (1, trifluoroacetic acid, aqueous methanol, 45 °C, 82%).8 The H-2 and C-2 resonances (as well as all other resonances) of the bis(trifluoroacetate) salt of lomaiviticin B (2) obtained in this way were in exact agreement with literature values. The free base of 2 was obtained by a basic wash. The chemical shifts of H-2 and C-2 of the free base form were in better agreement with (-)-lomaiviticin aglycon (6, free base of 2: δ H-2 = 3.64, δ C-2 = 71.3, CD₃OD). Thus, the chemical shift discrepancies observed between (-)-lomaiviticin aglycon (6) and natural lomaiviticin B (2) are attributed to the charged state of **2** when it was obtained from the producing organism.^{1a}

Hydrodediazotization Studies. A growing body of data suggests the cytotoxic effects of the kinamycins and lomaiviticins are mediated by a reducing cofactor,⁵ and the dimeric and monomeric diazofluorene derivatives prepared above provide an opportunity to probe substituents effects on the redox activity of the diazofluorene. We evaluated the rate of reduction of the unnatural *anti,anti-aglycon* **75**, (–)-lomaiviticin aglycon (**6**), the monomeric lomaiviticin aglycon **72**, and (–)-kinamycin C (**8**) by dithiothreitol (1 equiv) in methanol at 37 °C (Table 6). The *anti,anti-*dimer **75** was studied because it contains the flexibility and conjugated ketone substituents found within lomaiviticin A (**1**). Under these conditions, the *anti,anti-*dimer **75** and the monomeric lomaiviticin aglycon (**72**) undergo hydrodediazotization to form the hydroxyfulvenes **99** and **100**, respectively, and more slowly, the products



75 DTT (1 e or 72 CH ₃ OH,	equiv) 37 °C HC	о он 99 о		но о + но он з 10	1 or 102	
			composi	ition $(\%)^a$		
time (h)	75	99	101	72	100	102
0	100	0	0	100	0	0
1	62	38	0	89	10	1
10	22	78	0	76	11	13
^a Determined	l by LIPI	C/MS	analysis			

of methanolysis (101 and 102).⁵¹ Interestingly, these data reveal that the rate of reduction of the anti,anti-dimer 75 is ca. 2-fold faster than that of the monomeric lomaiviticin aglycon (72). Perhaps more strikingly, under otherwise identical conditions, neither (-)-lomaiviticin aglycon (6) nor (-)-kinamycin C (8) formed detectable levels of reduction products (UPLC/MS analysis). Collectively, these data suggest that the ketone function of lomaiviticin A (1) raises the reduction potential of the diazofluorene, allowing for facile electron transfer. Additionally, a stabilizing donor-acceptor interaction between the hydroxyfulvene and the remaining diazofluorene may further promote reductive activation of dimers such as 75, and lomaivitic A(1). Consequently, although lomaivitic A (1) and B (2) share a good deal of homology, the additional flexibility present in the structure of lomaivitic n A (1) is expected to lead to higher rates of reductive activation, which may be of biological significance.

Identification of Simple Diazofluorenes as Potent Inhibitors of Ovarian Cancer Stem Cells. In parallel with our synthetic studies, we have conducted structure–function studies of synthetic diazofluorenes. One aspect of this research has focused on evaluating the activity of our synthetic analogues against cancer stem cells (CSCs), a small population of cells within tumors that are highly tumorigenic and chemoresistant. CSCs have been identified in many different cancers, including ovarian cancer.⁴³

We have found that the diazofluorene **103** (Figure 4) potently inhibits the growth of cultured ovarian cancer stem



Figure 4. Structure of the anti-cancer stem cell agent 103.

cells (IC₅₀ = 500 nM). This compound is readily prepared (500 mg of **103** was synthesized in less than 1 week in our laboratory).⁸ Although the development of **103** as a clinical agent is of interest, of equal or greater significance are identification of the biological mechanisms underlying the cytotoxic effects of **103** in this cell line, an avenue of research we are currently pursuing.

CONCLUSION

We have described the evolution of enantioselective synthetic routes to (-)-kinamycin F (9) and (-)-lomaiviticin aglycon (6). Our synthetic routes were enabled by the development of scalable, efficient, and enantioselective syntheses of the

kinamycin and lomaiviticin D-ring precursors 60 and 68, and implementation of a convergent, three-step strategy for construction of the diazofluorene functional group. Use of substrate control to effect the transformation of the ketone 63 to the trans-1,2-diol 67, in the presence of the redox-sensitive diazofluorene, allowed for a short synthesis of (-)-kinamycin F (9, 13 steps from *m*-cresol).

Extensive investigations into the oxidative coupling of monomeric diazofluorenes such as 87 has allowed for a short (11-step) synthesis of (-)-lomaiviticin aglycon (6). These studies have revealed a complex relationship between substrate structure and reaction conditions, and stereochemical outcome and product distributions. Ultimately, conditions were found to form a precursor to lomaiviticin aglycon (6) in good yield (26-33%), and this was successfully advanced to the target.

Several new reactions, including a novel γ -quinonylation of β trimethylsilyl- α , β -unsaturated ketones, and a new α -quinonylation of enoxysilanes, were developed. In addition, we have described the application of nonaflylazide as a storable, benchstable surrogate for the volatile and highly reactive reagent trifluoromethanesulfonyl azide. We have also described the first use of manganese tris(hexafluoroacetylacetonate) (94) in the oxidative coupling of enoxysilanes. This reagent possesses physical properties (high solubility in nonpolar solvents and a coordinatively saturated metal center) that are complementary to more common oxidants (such as CAN), and may find applications in settings beyond those described here. We have obtained evidence that the flexible, dimeric structure of lomaiviticin A (1) enhances the rate of reductive activation of the metabolite, and we have identified simple diazofluorenes (103) as potent inhibitors of ovarian cancer stem cells.

The results reported herein lay the foundation for synthesis of lomaivitic A(1) itself. We have delineated efficient routes to advanced monomeric diazofluorenes and established their stability and reactivity profiles. The successful oxidative coupling of these systems provides encouragement that the dimerization of fully functionalized (glycosylated) monomeric diazofluorenes may be possible.

ASSOCIATED CONTENT

Supporting Information

Experimental procedures and detailed characterization data of all new compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

AUTHOR INFORMATION

Corresponding Author

seth.herzon@yale.edu

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

Financial support from the National Institute of General Medical Sciences (R01GM090000), the National Science Foundation (Graduate Research Fellowship to C.M.W.), the Searle Scholars Program, and Yale University is gratefully acknowledged. We thank Dr. Gil Mor and Dr. Ayesha Alvero for determining the activity of 103 against ovarian CSCs and Dr. Eric Paulson of the Yale Chemical and Biophysical Instrumentation Center for assistance with NMR. We acknowledge the Developmental Therapeutics Program of the National Cancer Institute for a gift of (-)-kinamycin C (8, NSC

138425). S.B.H. is a fellow of the David and Lucile Packard and the Alfred P. Sloan Foundations, and is a Camille Dreyfus Teacher-Scholar.

REFERENCES

(1) (a) He, H.; Ding, W. D.; Bernan, V. S.; Richardson, A. D.; Ireland, C. M.; Greenstein, M.; Ellestad, G. A.; Carter, G. T. J. Am. Chem. Soc. 2001, 123, 5362. (b) Woo, C. M.; Beizer, N. E.; Janso, J. E.; Herzon, S. B. J. Am. Chem. Soc. 2012, 134, 15285. For reviews, see: (c) Gould, S. J. Chem. Rev. 1997, 97, 2499. (d) Marco-Contelles, J.; Molina, M. T. Curr. Org. Chem. 2003, 7, 1433. (e) Arya, D. P. Top. Heterocycl. Chem. 2006, 2, 129. (f) Nawrat, C. C.; Moody, C. J. Nat. Prod. Rep. 2011, 28, 1426. (g) Herzon, S. B.; Woo, C. M. Nat. Prod. Rep. 2012, 29, 87.

(2) (a) Ito, S.; Matsuya, T.; Omura, S.; Otani, M.; Nakagawa, A. J. Antibiot. 1970, 23, 315. (b) Hata, T.; Omura, S.; Iwai, Y.; Nakagawa, A.; Otani, M. J. Antibiot. 1971, 24, 353. (c) Omura, S.; Nakagawa, A.; Yamada, H.; Hata, T.; Furusaki, A.; Watanabe, T. Chem. Pharm. Bull. 1971, 19, 2428. (d) Furusaki, A.; Matsui, M.; Watanabe, T.; Ōmura, S.; Nakagawa, A.; Hata, T. Isr. J. Chem. 1972, 10, 173. (e) Cone, M. C.; Seaton, P. J.; Halley, K. A.; Gould, S. J. J. Antibiot. 1989, 42, 179. (f) Seaton, P. J.; Gould, S. J. J. Antibiot. 1989, 42, 189.

(3) (a) Lin, H.-C.; Chang, S.-C.; Wang, N.-L.; Chang, L.-R. J. Antibiot. 1994, 47, 675. (b) Lin, H.-C.; Chang, S.-C.; Wang, N.-L.; Chang, L.-R. J. Antibiot. 1994, 47, 681.

(4) (a) Moore, H. W. Science 1977, 197, 527. For a review, see: (b) Rockwell, S.; Sartorelli, A. C.; Tomasz, M.; Kennedy, K. A. Cancer Metastasis Rev. 1993, 12, 165.

(5) (a) Arya, D. P.; Jebaratnam, D. J. J. Org. Chem. 1995, 60, 3268. (b) Laufer, R. S.; Dmitrienko, G. I. J. Am. Chem. Soc. 2002, 124, 1854.

(c) Feldman, K. S.; Eastman, K. J. J. Am. Chem. Soc. 2005, 127, 15344.

(d) Feldman, K. S.; Eastman, K. J. J. Am. Chem. Soc. 2006, 128, 12562. (e) Zeng, W.; Ballard, T. E.; Tkachenko, A. G.; Burns, V. A.; Feldheim, D. L.; Melander, C. Bioorg. Med. Chem. Lett. 2006, 16, 5148. (f) Hasinoff, B. B.; Wu, X.; Yalowich, J. C.; Goodfellow, V.; Laufer, R. S.; Adedayo, O.; Dmitrienko, G. I. Anti-Cancer Drugs 2006, 17, 825. (g) O'Hara, K. A.; Wu, X.; Patel, D.; Liang, H.; Yalowich, J. C.; Chen, N.; Goodfellow, V.; Adedayo, O.; Dmitrienko, G. I.; Hasinoff, B. B. Free Radic. Biol. Med. 2007, 43, 1132. (h) Ballard, T. E.; Melander, C. Tetrahedron Lett. 2008, 49, 3157. (i) Khdour, O.; Skibo, E. B. Org. Biomol. Chem. 2009, 7, 2140. (j) Heinecke, C. L.; Melander, C. Tetrahedron Lett. 2010, 51, 1455. (k) O'Hara, K. A.; Dmitrienko, G. I.; Hasinoff, B. B. Chem. Biol. Interact. 2010, 184, 396. (1) Mulcahy, S. P.; Woo, C. M.; Ding, W. D.; Ellestad, G. A.; Herzon, S. B. Chem. Sci. 2012, 3, 1070. See also refs 1a,1b.

(6) (a) Lei, X.; Porco, J. A. J. Am. Chem. Soc. 2006, 128, 14790. (b) Kumamoto, T.; Kitani, Y.; Tsuchiya, H.; Yamaguchi, K.; Seki, H.; Ishikawa, T. Tetrahedron 2007, 63, 5189. (c) Nicolaou, K. C.; Li, H.; Nold, A. L.; Pappo, D.; Lenzen, A. J. Am. Chem. Soc. 2007, 129, 10356. (d) Woo, C. M.; Lu, L.; Gholap, S. L.; Smith, D. R.; Herzon, S. B. J. Am. Chem. Soc. 2010, 132, 2540. (e) Scully, S. S.; Porco, J. A. Angew. Chem., Int. Ed. 2011, 50, 9722. For studies toward the kinamycins, see: (f) Chen, N.; Carriere, M. B.; Laufer, R. S.; Taylor, N. J.; Dmitrienko, G. I. Org. Lett. 2008, 10, 381. (g) Scully, S. S.; Porco, J. A. Org. Lett. 2012, 14, 2646.

(7) (a) Nicolaou, K. C.; Denton, R. M.; Lenzen, A.; Edmonds, D. J.; Li, A.; Milburn, R. R.; Harrison, S. T. Angew. Chem., Int. Ed. 2006, 45, 2076. (b) Krygowski, E. S.; Murphy-Benenato, K.; Shair, M. D. Angew. Chem., Int. Ed. 2008, 47, 1680. (c) Morris, W. J.; Shair, M. D. Org. Lett. 2008, 11, 9. (d) Zhang, W.; Baranczak, A.; Sulikowski, G. A. Org. Lett. 2008, 10, 1939. (e) Gholap, S. L.; Woo, C. M.; Ravikumar, P. C.; Herzon, S. B. Org. Lett. 2009, 11, 4322. (f) Nicolaou, K. C.; Nold, A. L.; Li, H. Angew. Chem., Int. Ed. 2009, 48, 5860. (g) Lee, H. G.; Ahn, J. Y.; Lee, A. S.; Shair, M. D. Chem.-Eur. J. 2010, 16, 13058. (h) Morris, W. J.; Shair, M. D. Tetrahedron Lett. 2010, 51, 4310. (i) Herzon, S. B.; Lu, L.; Woo, C. M.; Gholap, S. L. J. Am. Chem. Soc. 2011, 133, 7260. (j) Baranczak, A.; Sulikowski, G. A. Org. Lett. 2012, 14, 1027.

(8) See Supporting Information.

Journal of the American Chemical Society

(9) Díez, E.; Fernández, R.; Gasch, C.; Lassaletta, J. M.; Llera, J. M.; Martín-Zamora, E.; Vázquez, J. *J. Org. Chem.* **1997**, *62*, 5144.

(10) (a) Middleton, W. J. Tris(substituted amino)sulfonium Salts. U.S. Patent 3,940,402, 1976. (b) Noyori, R.; Nishida, I.; Sakata, J. *Tetrahedron Lett.* **1980**, 21, 2085. (c) Noyori, R.; Nishida, I.; Sakata, J.; Nishizawa, M. J. Am. Chem. Soc. **1980**, 102, 1223.

(11) Huot, R.; Brassard, P. Can. J. Chem. 1974, 52, 838.

(12) (a) Tamayo, N.; Echavarren, A. M.; Paredes, M. C. J. Org. Chem.

1991, 56, 6488. (b) Liebeskind, L. S.; Riesinger, S. W. J. Org. Chem. 1993, 58, 408.

(13) Tietze, L. F.; Gericke, K. M.; Schuberth, I. Eur. J. Org. Chem. 2007, 2007, 4563.

(14) Fujiwara, Y.; Domingo, V.; Seiple, I. B.; Gianatassio, R.; Del Bel, M.; Baran, P. S. J. Am. Chem. Soc. 2011, 133, 3292.

(15) (a) Ito, Y.; Hirao, T.; Saegusa, T. J. Org. Chem. 1978, 43, 1011.
(b) Larock, R. C.; Hightower, T. R.; Kraus, G. A.; Hahn, P.; Zheng, D. Tetrahedron Lett. 1995, 36, 2423.

(16) (a) Reich, H. J.; Reich, I. L.; Renga, J. M. J. Am. Chem. Soc. **1973**, 95, 5813. (b) Sharpless, K. B.; Launer, R. F.; Teranishi, A. Y. J. Am. Chem. Soc. **1973**, 95, 6137.

(17) Pilcher, A. S.; DeShong, P. J. Org. Chem. 1996, 61, 6901.

(18) (a) Fiallo, M. M. L.; Garnier-Suillerot, A. Biochemistry 1986, 25, 924. (b) Fiallo, M. M. L.; Garnier-Suillerot, A. Inorg. Chim. Acta 1987,

137, 119.

(19) Doering, W. v. E.; DePuy, C. H. J. Am. Chem. Soc. 1953, 75, 5955.

(20) Taber, D. F.; Ruckle, R. E.; Hennessy, M. J. J. Org. Chem. 1986, 51, 4077.

(21) Baum, J. S.; Shook, B. C.; Davies, H. M. L.; Smith, H. D. Synth. Commun. 1987, 17, 1709.

(22) Goddard-Borger, E. D.; Stick, R. V. Org. Lett. 2007, 9, 3797.

(23) Cavender, C. J.; Shiner, V. J. J. Org. Chem. 1972, 37, 3567.

(24) Zhu, S.-Z. J. Chem. Soc., Perkin Trans. 1 1994, 2077.

(25) Kolb, H. C.; VanNieuwenhze, M. S.; Sharpless, K. B. Chem. Rev. **1994**, *94*, 2483.

(26) Evans, D. A.; Chapman, K. T.; Carreira, E. M. J. Am. Chem. Soc. 1988, 110, 3560.

(27) Brown, H. C.; Vogel, F. G. M. Liebigs Ann. Chem. 1978, 695.

(28) Hawkins, E. G. E.; Large, R. J. Chem. Soc., Perkin Trans. 1 1974, 280.

(29) (a) Ito, Y.; Konoike, T.; Harada, T.; Saegusa, T. J. Am. Chem. Soc. 1977, 99, 1487. (b) Baran, P. S.; DeMartino, M. P. Angew. Chem., Int. Ed. 2006, 45, 7083. (c) DeMartino, M. P.; Chen, K.; Baran, P. S. J. Am. Chem. Soc. 2008, 130, 11546.

(30) (a) Baciocchi, E.; Casu, A.; Ruzziconi, R. *Tetrahedron Lett.* **1989**, 30, 3707. (b) Avetta, C. T.; Konkol, L. C.; Taylor, C. N.; Dugan, K. C.; Stern, C. L.; Thomson, R. J. *Org. Lett.* **2008**, *10*, 5621.

(31) Narasaka, K.; Okauchi, T.; Tanaka, K.; Murakami, M. Chem. Lett. **1992**, *21*, 2099.

(32) Ji, N.; O'Dowd, H.; Rosen, B. M.; Myers, A. G. J. Am. Chem. Soc. 2006, 128, 14825.

(33) This compound was previously prepared by Nicolaou and coworkers (ref 7f), and was named "the monomeric unit of lomaiviticin aglycon". Spectroscopic data for 72 were in agreement with the material synthesized by Nicolaou and co-workers.

(34) (a) Surya Prakash, G. K.; Hu, J.; Olah, G. A. *J. Fluorine Chem.* **2001**, *112*, 355. (b) Hata, H.; Kobayashi, T.; Amii, H.; Uneyama, K.; Welch, J. T. *Tetrahedron Lett.* **2002**, *43*, 6099.

(35) (a) Wender, P. A.; Hilinski, M. K.; Mayweg, A. V. W. Org. Lett. 2004, 7, 79. (b) Chen, K.; Baran, P. S. Nature 2009, 459, 824.

(36) Ohnemüller, U. K.; Nising, C. F.; Encinas, A.; Bräse, S. Synthesis 2007, 2175.

(37) Evans, S.; Hamnett, A.; Orchard, A. F.; Lloyd, D. R. Faraday Discuss. Chem. Soc. **1972**, 54, 227.

(38) Bryant, J. R.; Taves, J. E.; Mayer, J. M. Inorg. Chem. 2002, 41, 2769.

(39) Connelly, N. G.; Geiger, W. E. Chem. Rev. 1996, 96, 877.

(40) Schmittel, M.; Burghart, A. Angew. Chem., Int. Ed. 1997, 36, 2550.

(41) Bockman, T. M.; Kochi, J. K. J. Chem. Soc., Perkin Trans. 2 1996, 1633.

(42) Myers, A. G.; Fundy, M. A. M.; Lindstrom, P. A., Jr. Tetrahedron Lett. 1988, 29, 5609.

(43) (a) Zhou, J. B.; Zhang, Y. Exp. Opin. Drug. Discov. 2009, 4, 741.
(b) Alvero, A. B.; Chen, R.; Fu, H.-H.; Montagna, M.; Schwartz, P. E.; Rutherford, T.; Silasi, D.-A.; Steffensen, K. D.; Waldstrom, M.; Visintin, I.; Mor, G. Cell Cycle 2009, 8, 158. (c) Bapat, S. A. Reproduction 2010, 140, 33. (d) Wei, X.; Dombkowski, D.; Meirelles, K.; Pieretti-Vanmarcke, R.; Szotek, P. P.; Chang, H. L.; Preffer, F. I.; Mueller, P. R.; Teixeira, J.; MacLaughlin, D. T.; Donahoe, P. K. Proc. Natl. Acad. Sci. U.S.A. 2010, 107, 18874.